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Stereocontrolled and Convergent Entry to CF_2 -Sialosides: Synthesis of CF_2 -Linked Ganglioside GM4

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Gangliosides (sialic acid-containing glycosphingolipids) are known to be involved in intra- and intercellular cell signaling.^{1,2} For example, GM3 binds to epidermal growth factor receptor (EGFR) and inhibits EGFR-dependent cell proliferation.³ Among the four known human sialidases (neuraminidase),⁴ NEU2, NEU3, and NEU4 but not NEU1 cleave the glycosidic linkage of sialic acid in gangliosides.4a,5 Plasma membrane-associated sialidase (NEU3) seems to play a critical role in cell survival because it is upregulated in human cancer cells and tissues.⁶ However, the physiological roles of gangliosides are still not fully clarified, partly because rapid metabolism of gangliosides makes biological research difficult. For example, sialidases hydrolyze GM3 and GM4 (1) to lactosylceramide and galactosylceramide,⁷ which show different biological activities.⁸ Therefore, chemically and biologically stable ganglioside analogues would be very useful as probes for research to elucidate the roles of these gangliosides in normal and cancer cells. α-Difluorophosphonate derivatives are excellent nonhydrolyzable phosphate ester mimics, and the difluoromethylene group is now recognized as bioisosteric to an oxygen atom.⁹ Therefore, we envisioned the synthesis of difluoromethylene-linked (CF2linked) $\alpha(2,3)$ sially galactose (Figure 1) as a core structure of sialidase-resistant ganglioside mimics,¹⁰ because the $\alpha(2,3)$ sialylgalactose structure is a key structure of not only GM3 and GM4, but also most other gangliosides.

Various C-glycoside compounds, in which the oxygen atom of the glycosidic linkage is replaced by a carbon atom, have already been synthesized as hydrolytically stable glycoside mimics,¹¹ but efficient and stereoselective synthesis of C-sialoside is difficult, because the anomeric position of the sialoside (C2') is a tetrasubstituted carbon center.¹² To our knowledge, only two methodologies for the stereoselective synthesis of the C-linked $\alpha(2,3)$ sialylgalactose have been developed,¹³ and synthesis of CF₂-linked $\alpha(2,3)$ sially galactose has not been reported. Linhardt and co-workers have reported a convergent and stereoselective synthesis of CH-(OH)-linked $\alpha(2,3)$ sially galactose derivatives based on the SmI₂mediated coupling reaction^{13a,14} of the sialylsulfone derivative with the aldehyde prepared from galactose. First, we examined transformation of their CH(OH)-linked $\alpha(2,3)$ sially galactose to CF₂linked $\alpha(2,3)$ sialylgalactose. However, this was unsuccessful, probably due to the steric hindrance around the C-glycoside carbon linkage.¹⁵ Therefore, for the synthesis of the CF₂-linked $\alpha(2,3)$ sialylgalactose unit, a new strategy was required. Herein we report a convergent and highly stereoselective method for the construction of the CF₂-linked $\alpha(2,3)$ sialylgalactose unit, and the conversion of this unit to the CF_2 -linked analogue of ganglioside GM4 (2).¹⁶

As shown in Figure 2, we planned to construct the key C2'– CF₂ bond via Ireland–Claisen rearrangement of the ester III.¹⁷ We anticipated that rearrangement would occur from the α -face of the anomeric center (C2') through the chairlike transition state II after formation of the (Z)-silyl enolate. The resulting product I could be







Figure 2. Strategy for the stereocontrolled synthesis of CF2-sialoside.

converted to the desired CF_2 - $\alpha(2,3)$ sialylgalactose unit **3** in a stereocontrolled manner.

The precursor for the Ireland–Claisen rearrangement **9** was prepared by condensation of **4**¹⁸ with **8**, which was prepared from **6**¹⁵ as shown in Scheme 1. Ireland–Claisen rearrangement of **9** proceeded smoothly even at ambient temperature on treatment with LHMDS and TMSCl in THF. After treatment with TMS– diazomethane, methyl ester **10** possessing a α -*CF*₂-sialoside linkage was obtained as a single isomer in 86% yield. The strong HMBC correlation between α 3'-H and 1'-C indicated that the C2' tetrasubstituted carbon center has the desired α -stereochemistry.

To achieve stereocontrolled introduction of a C2 hydroxyl group into the galactose unit, 10 was converted to the conformationally fixed lactone 11. After removal of *p*-methoxybenzylidene acetal, protection of the 6-OH group with TBS, and saponification of the methyl ester, the desired lactone 11 was obtained in good yield by treatment with carbodiimide (Scheme 2). Although attempts at hydroboration or epoxidation of 11 were unsuccessful, we were pleased to find that dihydroxylation of 11 using a stoichiometric amount of OsO4 proceeded in a completely stereoselective manner to afford the desired diol 12. Regio- and stereoselective reduction of the C3-hydroxyl group was successfully achieved by radical reduction of the cyclic thiocarbonate. Namely, treatment of 12 with thiophosgene, followed by AIBN and Bu₃SnH, gave 13 as a single isomer. Hydrogenolysis of the four BOM groups, together with removal of the TBS group, afforded the key 4-methoxyphenyl CF2linked $\alpha(2,3)$ sialylgalactose lactone unit 14. The stereochemical assignment of the newly formed chiral carbon centers (C2', C2, and C3) was confirmed by X-ray crystallographic analysis of 14.

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Scheme 1^a



^a Reaction conditions: (a) CF₂Br₂, HMPT, THF, rt, 4 h, 76% (see ref 19); (b) conc. HCl, MeOH-CH2Cl2 (4:1), rt, 92%; (c) (4-OMe)Ph-CH(OMe)₂, TsOH, CH₃CN, rt; (d) TBAF, THF, rt, 74% (two steps); (e) EDC·HCl, DMAP, 4, CH₂Cl₂, rt, 72%; (f) LHMDS, TMSCl, THF, -78 °C then rt; (g) TMSCHN₂, Et₂O-MeOH (1:1), 86% (two steps).

Scheme 2^a



 a Reaction conditions: (a) 80% AcOH aq., rt, 80%; (b) TBSCl, Et_3N, DMAP, CH_2Cl_2, rt, 89%; (c) 2 M KOH aq., THF, 60 °C; (d) EDC+HCl, DMAP, CH₂Cl₂, rt, 77% (two steps); (e) OsO₄ (1.7 equiv), pyridine, rt, then sat. NaHSO3 aq., rt, 90%; (f) thiophosgene, DMAP, CH2Cl2, rt; (g) Bu₃SnH, AIBN, toluene, 100 °C, 84% (two steps); (h) Pd(OH)₂/C, H₂, MeOH, rt, 99%; (i) Ac₂O, pyridine, rt, 94%; (j) Ce(NH₄)₂(NO₃)₆, CH₃CN-H₂O (4:1), rt; (k) Cl₃CCN, DBU, CH₂Cl₂, rt, 75% (two steps); (l) 16, TfOH, CH2Cl2, 0 °C, 33% (based on recovery); (m) NaOMe, MeOH, rt, then H2O, rt 60%

The overall yield of this CF_2 -linked sialoside unit from 6 was 13% (15 steps).

To demonstrate the potential of this novel CF_2 -linked sialylgalactose unit 14 as an intermediate for the synthesis of ganglioside analogues, conversion of 14 to CF_2 -linked GM4 (2) was performed. After acetylation of all hydroxyl groups, the MP group at the anomeric position was converted to trichloroacetoimidate to give a glycosyl donor 15. Glycosylation of 15 with the ceramide derivative 16^{20} was conducted in the presence of TfOH in CH₂Cl₂. Finally, synthesis of 2 was completed by methanolysis and hydrolysis. Although the yield of glycosidation with ceramide needs to be improved, to our knowledge, this is the first synthesis of a ganglioside analogue containing a CF_2 -sialoside linkage.

 CF_2 -linked GM4 (2) showed moderate inhibition of NEU2 (IC₅₀ = 754 μ M) and NEU4 (IC₅₀ = 930 μ M).¹⁵ A preliminary study indicated that 2 also showed remarkable inhibition for human lymphocyte proliferation.^{15,21} These results suggest that CF_2 sialoside can indeed act as a mimic of O-sialoside.

In conclusion, the CF_2 - $\alpha(2,3)$ sialy galactose unit was synthesized in a completely stereoselective manner, and this nonhydrolyzable

sialylgalactose unit was confirmed to be suitable as a glycosyl donor for the synthesis of a GM4 analogue. This new strategy based on Ireland-Claisen rearrangement should be applicable not only for the synthesis of CF_2 -sialylgalactose but also for the construction of various other (un)substituted C-sialoside units, which are expected to be useful for the synthesis of novel gangliosidemimicking molecules. Synthesis of larger gangliosides, such as a GM3, and biological experiments using CF_2 -linked ganglioside analogues are currently under way.

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Supporting Information Available: Experimental procedure, characterization of new compounds, biological evaluations, ¹H NMR and ¹³C NMR spectra, and CIF file for compound 14. This material is available free of charge via the Internet at http://pubs.acs.org.

References

- (a) Hakomori, S. Proc. Natl. Acad. Sci. U.S.A. 2002, 99, 10231. (b) (1)Sonnino, S.; Mauri, L.; Chigorno, V.; Prinetti, A. Glycobiology 2007, 17, 1R
- (a) Hakomori, S. *Proc. Natl. Acad. Sci. U.S.A.* **2002**, *99*, 225. (b) Yoon, S.-J.; Nakayama, K.; Hikita, T.; Handa, K.; Hakomori, S. *Proc. Natl. Acad.* (2)Sci. U.S.A. 2006, 103, 18987.
- (a) Rebbaa, A.; Hurh, J.; Yamamoto, H.; Kersey, D. S.; Bremer, E. G. *Glycobiology* **1996**, *6*, 399. (b) Miljan, E. A.; Meuillet, E. J.; Mania-(3)Farnell, B.; George, D.; Yamamoto, H.; Simon, H.-G.; Bremer, E. G. J. Biol. Chem. 2002, 277, 10108.
- (a) Miyagi, T.; Wada, T.; Yamaguchi, K.; Hata, K. Glycoconjugate J. (4)2004, 20, 189 and references therein. (b) Monti, E.; Bassi, M. T.; Bresciani, R.; Civini, S.; Croci, G. L.; Papini, N.; Riboni, M.; Zanchetti, G.; Ballabio, A.; Preti, A.; Tettamanti, G.; Venerando, B.; Borsani, G. *Genomics* 2004, 83, 445. (c) Yamaguchi, K.; Hata, K.; Koseki, K.; Shiozaki, K.; Akita, H.; Wada, T.; Moriya, S.; Miyagi, T. *Biochem. J.* 2005, 390, 85.
 (5) Seyrantepe, V.; Landry, K.; Trudel, S.; Hassan, J. A.; Morales, C. R.;
- Pshezhetsky, A. V. J. Biol. Chem. 2004, 279, 37021.
- Wada, T.; Hata, K.; Yamaguchi, K.; Shiozaki, K.; Koseki, K.; Moriya,
- S.; Miyagi, T. Oncogene 2007, 26, 2483.
 (a) Wada, T.; Yoshikawa, Y.; Tokuyama, S.; Kuwabara, M.; Akita, H.; Miyagi, T. Biochem. Biophys. Res. Commun. 1999, 261, 21. (b) Oehler, C.; Kopitz, J.; Cantz, M. Biol. Chem. 2002, 383, 1735.
- (8) (a) Ueno, S.; Saito, S.; Wada, T.; Yamaguchi, K.; Satoh, M.; Arai, Y.; Miyagi, T. J. Biol. Chem. 2006, 281, 7756. (b) Coetzee, T.; Fujita, N. Dupree, J.; Shi, R.; Blight, A.; Suzuki, K.; Suzuki, K.; Popko, B. Cell **1996**, 86, 209.
- (9) Burke, T. R., Jr. *Curr. Top. Med. Chem.* 2006, 6, 1465.
 (10) For *CF*₂-linked *disaccharides*: (a) Herpin, T. F.; Motherwell, W. B.; Tozer, M. J. *Tetrahedron: Asymmetry* 1994, 5, 2269. (b) Kovensky, J.; Burrieza, D.; Colliou, V.; Cirelli, A. F.; Sinay, P. J. Carbohydr. Chem. 2000, 19, 1. (c) Berber, H.; Brigaud, T.; Lefebvre, O.; Plantier-Royon, R.; Portella, C. Chem. Eur. J. 2001, 7, 903. (d) Tony, K. A.; Denton, R. W.; Dilhas, A.; Jiménez-Barbero, J.; Mootoo, D. R. Org. Lett. 2007, 9, 1441.
 Beau, J.-M.; Vauzeilles, B.; Skrydstrup, T. C-Oligosaccharide Synthesis. In Glycoscience: Chemistry and Chemical Biology; Fraiser-Reid, B. O.,
- Tatsuta, K., Thiem, J., Eds.; Springer: Heidelberg, Germany, 2001; Vol. a) p 2679.
 (12) (a) Wallimann, K.; Vasella, A. *Helv. Chim. Acta* 1991, 74, 1520. (b) Nagy,
- J. O.; Bednarski, M. D. *Tetrahedron Lett.* **1991**, *32*, 3953. (c) Paulsen, H.; Matschulat, P. *Liebigs. Ann. Chem.* **1991**, 487.
 (13) (a) Vlahov, I. R.; Vlahova, P. I.; Linhardt, R. J. J. Am. Chem. Soc. **1997**,
- 119, 1480. (b) Notz, W.; Hartel, C.; Waldscheck, B.; Schmidt, R. R. J. Org. Chem. 2001, 66, 4250.
- (14) Malapelle, A.; Abdallah, Z.; Doisneau, G.; Beau, J.-M. Angew. Chem., Int. Ed. 2006, 45, 6016-6020.
- (15) See Supporting Information. (16) *CH(OH)*-linked and *CH*₂-linked Sia α (2,6)-GalNAc derivatives: (a) Kuberan, B.; Sikkander, S. A.; Tomiyama, H.; Linhardt, R. J. Angew. Chem., Int. Ed. 2003, 42, 2073. (b) Abdallah, Z.; Doisneau, G.; Beau, J.-M. Angew. Chem., Int. Ed. 2003, 42, 5209
- (a) Ireland, R. E.; Mueller, R. H.; Willard, A. K. J. Am. Chem. Soc. 1976, 98, 2868.
 (b) Daman, D. B.; Hoover, D. J. J. Am. Chem. Soc. 1990, 112, 6439.
 (c) Patel, S. T.; Percy, J. M.; Wilkes, R. D. Tetrahedron 1995, 51, 11327.
 (d) Recently non-selective Ireland-Craisen rearrangement of a (17)uronic acid derivative was reported: Werschkun, B.; Thiem, J. Tetrahedron: Asymmetry 2005, 16, 569.
- (18) Schmid, W.; Christian, R.; Zbiral, E. Tetrahedron Lett. 1988, 29, 3643.
- (19) Zhao, Z.; Liu, H. J. Org. Chem. 2001, 66, 6810.
 (20) Schmidt, R. R.; Zimmermann, P. Tetrahedron Lett. 1986, 27, 481.
- (21) Ladisch, S.; Hasegawa, A.; Li, R.; Kiso, M. Biochemistry 1995, 34, 1197.

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